Using forage litter to improve soil degradation of pure Betula platyphylla forest in the Loess Plateau, China

Luc Nhu Trung¹,², Zengwen Liu³,⁴*, Xiaoxi Zhang⁵, Yuanhao Bing⁶ and Bochao Zhu⁷


Abstract. The long-term growth of pure forest is an important issue that affects stability and sustainable development of ecosystem, while using forage litter as fertilizer or directly establishing tree-grass complex vegetation may be the most effective prevention way. This study took the artificial pure forests of Betula platyphylla, which were widely distributed in the Loess Plateau of China, as the object and conducted a 120-day decomposition incubation experiment of forest humus soil mixed with seven common leguminous forage litters to study the effects of forage litters in controlling the degradation of soil biological and chemical properties of pure forests. The results showed that: by adding forage litter to the soil of pure B. platyphylla forest, litters of Lespedeza bicolor and Onobrychis viciaefolia improved the soil quality obviously, followed by Astragalus adsurgens and Melilotus oficinalis, while Medicago sativa lead to obvious deterioration, followed by Vicia villosa and Coronilla varia.

Key words: forage litter, Betula platyphylla, pure forest, soil degradation.

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Introduction

Artificial pure forest made of Betula platyphylla Sukaczew, one of the main afforestation of broad-leaved tree species in the Loess Plateau, is important vegetation type for comprehensive treatment of the Loess Plateau of China. However, it has been reported in large number of studies that after long-term growth or continuous planting for several generations, pure forest often results in soil deterioration (Joshi et al., 1997; Zhang & Cai, 2004). The reasons include pure forest’s extreme selection of absorbing and returning of soil nutrient, and the particular impact of the single tree species litter on soil biochemical properties, which cause the soil properties deviating from its original equilibrium to a negative (destroy) development. Previous studies indicated that, in the central hilly area of the Loess Plateau (took Huangling of Shaanxi province as an example), the pure forest of B. platyphylla had resulted in...
a huge loss of soil available P and K, and decrease of activities of enzymes such as sucrase and phosphatase, while the activity of protease was increased to some extent (Liu et al., 2007; Liu et al., 2009).

In terms of the soil degradation, establishing tree-grass complex vegetation is the most effective solution to prevent it, not only because of the rapid growth of grass and the wide range source of litter (Zhang et al., 2013), but also a large amount of forage (especially leguminous forage) can improve soil properties in terms of total N, available P and K contents, as well as the other soil biochemical properties (Li, 2008; Pan et al., 2011). According to the above assumption, in this study, humus soil in the pure forests of B. platyphylla were gathered and a 120-day decomposition incubation experiment of forest soils mixed with seven common leguminous forage litters were conducted to study the effects of forage litters on soil, in order to bring some scientific guidance for the prevention of soil degradation and the improvement of local pure forests.

### Material and Methods

#### Study sites

Soil from B. platyphylla pure forest plantations was collected from Huangling county of Shaanxi province, China (Table 1). This area is located in a typical hilly region of the Loess Plateau. This region is classified as semi-humid warm-temperate zone, with an average annual temperature of 9.4 °C. The average annual rainfall is 630.9 mm, the average frost-free period is approximately 150 days, and the relative humidity is approximately 64%. The soil is a typical grey cinnamon soil, which is classified as Ultisols in the USDA Soil Taxonomy system.

#### Collection of soil samples and forage litter

For this study, soil collections were made from B. platyphylla pure forests that had reached the mature growth stage. In these forests, the tree growth density was greater than 0.9, and the understory vegetation coverage was less than 15%. Standard 20 m × 20 m plots were established in typical locations in two forests, and the site factors and tree growth indexes were measured in these plots (Table 1). Five quadrats were established in the plots, each 1 m×1 m. The humus layer of the soil was collected at a depth of 0–10 cm after removing the litter from the ground. The samples from each stand were then mixed to form a composite sample and transported to the laboratory, then passed through 5 mm mesh sieves to get rid of leaves, roots, and gravel. Simultaneously, litters from Astragalus adsurgens Pall., Lespedeza bicolor Turcz., Vicia villosa Roth, Corinilla varia L., Melilotus officinalis (L.) Pall., Medicago sativa L. and Onobrychis viciefolia Scop. were collected in late autumn before withered, gently washed and oven-dried at 65 °C for 24 h to reach constant weight. The dry litter samples were then ground with a laboratory mill (φ = 1 mm).

#### Decomposition incubation of soil mixtures with forage litter

Prepared fresh soil samples and litter were mixed with a dry weight ratio of 100:2 (with the original forest soil with no litter as a control). A total of 2.5 kg mixed soil was placed in impermeable plastic pots with natural density. Each treatment had

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### Table 1. Introduction of pure forests.

<table>
<thead>
<tr>
<th>Pure forests</th>
<th>Physiognomy positions</th>
<th>Altitude (m)</th>
<th>Aspect</th>
<th>Slope (°)</th>
<th>Average age (years)</th>
<th>Density (/hm²)</th>
<th>BWD (cm)</th>
<th>Average height (m)</th>
</tr>
</thead>
<tbody>
<tr>
<td>B. platyphylla</td>
<td>Upper slope</td>
<td>1180</td>
<td>NE20°</td>
<td>25°</td>
<td>35</td>
<td>4400</td>
<td>11.41</td>
<td>9.8</td>
</tr>
</tbody>
</table>
3 replications. Then, a measured amount of distilled water was added to each pot to adjust the soil moisture to 50% of saturated field capacity, pots were covered with plastic film with 4 holes for ventilation to reduce water loss, and incubated at room temperature (20–25 °C). The pots were weighed every three days and distilled water was added with a sprayer to keep the weight of pots constant (so that the moisture was constant as well). Samples of mixed soil were incubated under these conditions for 120 days until inner litter decomposed and no residual visible to naked eye. After incubation, soil samples were placed on clean plates, and the remaining leaf litter was carefully removed. A part of soil was collected to measure the content of micro-organisms. The remaining soil samples were air-dried, and ground for the measurement of following biological and chemical properties.

**Measurements of soil properties**

Chemical properties: The chemical properties of the soil were measured according to methods suggested by Lu (1999). Soil pH was measured by glass electrode method (the ratio of soil to water was 1:2.5). Organic matter was determined with the potassium bichromate titrimetric method. Available nitrogen was measured with the micro-diffusion technique. Available phosphorus was measured by NaHCO₃ extraction with the molybdenum blue colorimetric method. Available potassium was measured with the ammonium acetate extraction flame photometric method. Cation exchange capacity was measured with the sodium acetate-ammonium acetate flame photometric method.

Biological properties: The quantity of microorganisms was measured with the dilution plating method (Nanjing Institute of Soil Science..., 1985) (bacteria-beef extract peptone agar culture medium, fungi-potato dextrose agar culture medium, actinomyces-GAO 1st synthetic culture medium). Urease activity was measured with the phenol sodium-sodium hypochlorite colorimetric method. Sucrase activity was measured with the Na₂S₂O₃ titration method. Catalase activity was measured with the KMnO₄ titration method. Dehydrogenase activity was measured with the triphenyltetrazolium chloride colorimetric method. Alkaline phosphatase activity was measured with the disodium phenyl phosphate colorimetry method. Protease activity was measured with the ninhydrin colorimetric method. Polyphenol oxidase activity was measured with the iodometric titration method (Guan, 1986).

**Statistical analysis**

The data were analyzed with Microsoft Excel 2010 and SPSS 19.0 software. The significance of the differences between the effects of the forage litter types on the pure forest soil properties was evaluated with LSD test (α = 0.05). For the addition of forage litter had complex effects on the 16 indexes of soil biochemistry, while even the same litter affected the soil properties at different extents and directions. Thus to assess the comprehensive effects of forage litters on soil properties, a comprehensive principal component analysis method was employed to analyze the rate of improvement of 15 biochemical properties (pH excluded) relative to the CK values of the control sample (soil without any litter mixed). This analysis was performed with SPSS 19.0, and the comprehensive principal component value \( F_i \) was calculated (Figure 1). \( F_i > 0 \) implies that the addition of forage litter can improve soil properties (by decreasing the rate of negative polarization or increasing the rate of positive polarization); in contrast, \( F_i < 0 \) implies that the forage litter causes the soil properties to deteriorate.

**Results and Discussion**

According to the biochemical properties changed by adding different forage
litter to the soil of pure *B. platyphylla* forest (Table 2), litters of *A. adsurgens* significantly increased the content of organic matter, available N and K, the activity of sucrase and dehydrogenase, and activate the growth of fungi and actinomycetes, urease and catalase activity, and activate the growth of fungi and actinomycetes, but significantly reduced the activity of the urease, catalase and protease. Whereas *L. bicolor* significantly increased soil available K content, and activate the growth of fungi and actinomycetes, but significantly reduced the activity of the urease, phosphatase and polyphenol oxidase. On the other hand, *C. varia* significantly increased the soil available N, K contents, and the activity of sucrase, but meanwhile reduced the activity of the urease. An increased content of available N and K, enhanced activity of catalase and sucrase and active growth of fungi and actinomycetes were observed with *M. officinalis* treatment, but it significantly reduced the activity of the urease and protease. The effect of *M. sativa* was the same as *M. officinalis* except it significantly reduced the activity of polyphenol oxidase instead of protease. In contrary, *O. viciaefolia* significantly increased the content of available K, and the activity of the urease, sucrase and phosphatase, but significantly reduced the activity bacterial growth of catalase.

The comparison between the results of this study and the previous studies (the long-term growth of pure forest of *B. platyphylla* had resulted in a serious of degradation such as a huge loss of available N, P and K in soil and the decrease of sucrase, dehydrogenase, urease and phosphatase activity) showed that all forage litters reduced the loss of available K, in which the content of available P. All the forage could significantly mitigate the decrease of sucrase activity except *L. bicolor*, in which *M. sativa* showed the most obvious effect; *O. viciaefolia* could significantly mitigate the decrease of catalase activity, while *V. villosa* greatly exacerbated this trend.

Considering the impact of forage litter on the above degradation trends and the effect on other soil biochemical properties, according to the principal component analysis (Figure 1), after adding the forage litter to the soil of pure *B. platyphylla* forest, *L. bicolor* and *O. viciaefolia* showed obvious improvement effect, followed by *A. adsurgens* and *M. officinalis*; while *M. sativa* played an obvious deterioration role, followed by *V. villosa* and *C. varia*.

The previous studies showed that forage such as *A. adsurgens*, *M. sativa*, *L. bicolor* can improve the contents of total nitrogen and total phosphorus on the soil, among which, *M. sativa* associated with obvious improvement of total nitrogen, and *M. sativa* and *A. adsurgens* resulted in more obvious improvement of total phosphorus than *L. bicolor* (Zhang *et al*., 2008); *O. viciaefolia* and *M. sativa* improved organic matters and nitrogen contents of soil (Li, 2008); After *C. varia* was added to grey brown desert soil in Heihe valley, soil organic matters, available nitrogen, phosphorus and available potassium content were increased effectively (Wang & Jin, 2006). This may be due to rich nitrogen of legume forages, and low carbon nitrogen ratio which enhanced the soil microbial activity (Hong *et al*., 2003). Therefore, there was an improvement in the soil organic matters as well as the available phosphorus and rapidly available potassium (Wang *et al*., 2011b).

So, it is an effective means to rapidly improve the soil properties of pure forest and prevent the degradation of soil by means of quickly supplement of soil nutrient loss through the decomposition of other plant litter and the inducing the betterment of other biochemical properties.
Table 2. Chemical and biological properties of forest soil after mix-incubation for 120 days with different grass litter.

<table>
<thead>
<tr>
<th>Soil properties</th>
<th>Type of forage litter</th>
<th>Original soil of Betula platyphylla forest</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>A.a</td>
<td>L.b</td>
</tr>
<tr>
<td>Chemical proporties</td>
<td></td>
<td></td>
</tr>
<tr>
<td>pH</td>
<td>7.30±0.02 d</td>
<td>7.42±0.01 c</td>
</tr>
<tr>
<td>Org-M (g·kg⁻¹)</td>
<td>100.8±0.08 a</td>
<td>77.5±8±0.04 c</td>
</tr>
<tr>
<td>N Available N (mg·kg⁻¹)</td>
<td>526.4±6±30 d</td>
<td>423.5±6±30 e</td>
</tr>
<tr>
<td>P Available K (mg·kg⁻¹)</td>
<td>11.4±6±46 a</td>
<td>8.6±6±24 b</td>
</tr>
<tr>
<td>K Available N (mg·kg⁻¹)</td>
<td>572.0±6±57 d</td>
<td>484.2±6±19.47 f</td>
</tr>
<tr>
<td>CEC /(cmol kg⁻¹)</td>
<td>34.56±1.31 a</td>
<td>32.93±0.33 ab</td>
</tr>
<tr>
<td>Enzyme activity</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Urease (mg·kg⁻¹)</td>
<td>37.00±1.73 d</td>
<td>40.8±6±2.10 c</td>
</tr>
<tr>
<td>Sucrase (mL·g⁻¹)</td>
<td>2.2±6±0.14 c</td>
<td>1.6±6±0.06 c d</td>
</tr>
<tr>
<td>Catalase (mL·g⁻¹)</td>
<td>13.7±6±0.03 a</td>
<td>12.6±6±0.09 a</td>
</tr>
<tr>
<td>Dehydrogenase (mg·g⁻¹)</td>
<td>0.6±6±0.04 a</td>
<td>0.5±6±0.08 ab</td>
</tr>
<tr>
<td>Phosphatase (mg·kg⁻¹)</td>
<td>0.4±6±0.01 ab</td>
<td>0.4±6±0.03 ab</td>
</tr>
<tr>
<td>Protease (mg·kg⁻¹)</td>
<td>9.0±6±0.62 c</td>
<td>20.4±6±1.31 ab</td>
</tr>
<tr>
<td>Polyphenoloxidase (mL·g⁻¹)</td>
<td>0.0±6±0.000 a</td>
<td>0.0±6±0.002 a</td>
</tr>
<tr>
<td>Microorganism</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bacteria (10⁴·g⁻¹)</td>
<td>87.8±6±4.30 c</td>
<td>105.5±6±2.02 b</td>
</tr>
<tr>
<td>Fungi (10³·g⁻¹)</td>
<td>22.0±6±3.48 a</td>
<td>49.3±6±6.05 a</td>
</tr>
<tr>
<td>Actinomycetes (10⁴·g⁻¹)</td>
<td>21.8±6±6.33 a</td>
<td>10.9±6±4.22 b</td>
</tr>
</tbody>
</table>

Values sharing the same letters differ non-significantly (P > 0.05).

Note: A.a – Astragalus adsurgens, L.b – Leapedeza bicolor, V.v – Vicia villosa, C.v – Coronilla varia, M.o – Melilotus officinalis, M.s – Medicago sativa, O.v – Onobrychis vicieofolia. The value in table is the average ± SE.
This test showed that, in Huangling region, all forage litter could not significantly increase the content of available P and mitigate its degradation trend, which is not similar to the findings of Li et al. (2011a). An important reason for the above results may be that the decomposition of the litter itself and soil organic matter utilized a mass of the available P, because the decomposition of microbial requires appropriate C/P (Li et al., 2010; Zhao et al., 2011). The adding of all kinds of litter significantly alleviated the loss of available K or enhanced its increasing trend, which is similar to the findings of other scholars (Zhang et al., 2008). This is because that K is a non-structural element (Osono & Takeda, 2004), which is more easily to release compared with N and P (Zhao et al., 2011). The release of a large number of water-soluble K will increase the content of soil available K.

In terms of biological properties, adding variety of litters showed some improvement on enzyme activity, which is similar to the findings of Li (Liu et al., 2010; Li et al., 2011b; Wang et al., 2012). This might be caused due to enzymes released from litters itself, or the products of decomposition stimulate the activity of enzyme system. Some litters inhibited some activity, even intensifying its negative trend, which is similar to the findings of Sun et al. (2011). Because there are certain correlation between different properties of soil, after having been added in the soil, forage litter can make direct effect on some certain properties as well as make indirect effect on other properties (Li, 1993; Liu et al., 2004; Wang et al., 2011a). The above phenomenon may be due to that the enzyme was affected by the change of some nutrient elements (Sun et al., 2011). The changes in microbial activity was also subjected to multiple factors, such as the substrate quality of the litter (Qi et al., 2004), the content of special chemical substance, and the nutrient of the soil itself. After adding of forage litters the microbial species, the quantity and even the microbial flora (Kourtev et al., 2002) would change, which needs to be further investigated and explained.
Conclusions

All the forage litters increased the content of available K, in which the effect of C. variabilis was the most obvious. However, they could hardly control the loss of available P. All forage could improve sucrase activity except L. bicolor, and O. viciaefolia could improve catalase activity significantly.

The comprehensive analysis of results showed that after adding the forage litter to the soil of pure B. platyphylla forest, L. bicolor and O. viciaefolia got obvious improvement effects, followed by A. adscendens and M. officinalis, and these forage may be suitable to be planted with B. platyphylla for combating the degradation of soil properties. M. sativa played an obvious deterioration role, followed by V. villosa and C. variabilis, these forage should avoid to be planted with B. platyphylla.

References


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